

Berberine: A natural modulator of immune cells in multiple sclerosis

Esmaeil Yazdanpanah¹ | Sepehr Dadfar² | Alireza Shadab² | Niloufar Orooji² |
MohammadHossein Nemati² | Alireza Pazoki² | Seyed-Alireza Esmaili¹ |
Rasoul Baharlou^{2,3} | Dariush Haghmorad^{2,3} 

¹Immunology Research Center, Mashhad University of Medical Sciences, Mashhad, Iran

²Department of Immunology, School of Medicine, Semnan University of Medical Sciences, Semnan, Iran

³Cancer Research Center, Semnan University of Medical Sciences, Semnan, Iran

Correspondence

Dariush Haghmorad, Semnan University of Medical Sciences, Semnan, Iran.
Email: dhaghmorad@gmail.com

Abstract

Berberine is a benzyloquinoline alkaloid found in such plants as *Berberis vulgaris*, *Berberis aristata*, and others, revealing a variety of pharmacological properties as a result of interacting with different cellular and molecular targets. Recent studies have shown the immunomodulatory effects of Berberine which result from its impacts on immune cells and immune response mediators such as diverse T lymphocyte subsets, dendritic cells (DCs), and different inflammatory cytokines. Multiple sclerosis (MS) is a chronic disabling and neurodegenerative disease of the central nervous system (CNS) characterized by the recruitment of autoreactive T cells into the CNS causing demyelination, axonal damage, and oligodendrocyte loss. There have been considerable changes discovered in MS regards to the function and frequency of T cell subsets such as Th1 cells, Th17 cells, Th2 cells, Treg cells, and DCs. In the current research, we reviewed the outcomes of in vitro, experimental, and clinical investigations concerning the modulatory effects that Berberine provides on the function and numbers of T cell subsets and DCs, as well as important cytokines that are involved in MS.

KEYWORDS

berberine, dendritic cells, multiple sclerosis, Th1 and Th17 cells, Th2 and Treg cells

1 | INTRODUCTION

Multiple sclerosis (MS) is an autoimmune and neurodegenerative disease that affects about 2.5 million people in the world; a chronic neuroinflammatory disease that causes a disability in young people with an average age of 30 years, and women are at much higher risk of MS than men¹; the prevalence of this disease is 2 in 100,000 in

Japan, 100 in 100,000 in Northern Europe² and 29 in 100,000 in Iran.³ MS is one of the leading causes of disability in the United States.⁴

Despite the mechanism of affecting MS remains unknown, some genetics and environmental factors can increase susceptibility to this disease; for instance, being heterozygote for genetic locus HLA-DRB1*15:01 has a chance proportion of MS above 3 and homozygote of it

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2024 The Authors. *Immunity, Inflammation and Disease* published by John Wiley & Sons Ltd.

increases this ratio to above 6⁵; environmental factors including vitamin D deficiency, being a daily smoker and EBV infection. Symptoms of MS vary from central paralysis to difficulty walking, visual problems, fatigue, hearing loss, Amblyopia (lazy eyes), and even sexual impotence.^{3,6} The economy is negatively impacted by this disease because the ability to work in patients goes from 82% to 8%.⁷

The exact pathology of MS has not yet been discovered but based on multiple investigations, the main cause of MS is the infiltration of immune cells like T cells, B cells, and macrophage cells in the central nervous system (CNS) and starting inflammatory and immune reactions that lead to demyelination and axon loss in the white matter of the brain and spinal cord that disrupts the neuroaxonal signals^{1,5,8}; the main treatment for MS has not been discovered yet, but the base of the existing agents are: anti-inflammatory, immunomodulatory and immunosuppression drugs.^{1,8}

The immunopathology of MS can be explained by the study on mice in which MS is induced. In the experimental autoimmune encephalomyelitis (EAE), an animal model of MS, healthy mice are converted into MS mice in two different ways; first: injection of CNS antigens with immune activators,¹ second: adoptive transfer of activated T cells from EAE mice to healthy mice⁹; study on them has shown that various immune cells, such as T cells, B cells, macrophages, microglia, dendritic cells (DCs), monocytes, NKT cells, and other cells play an important role in the pathogenesis of the disease.^{10,11}

The main cells that lead to MS pathogenesis are adaptive immune cells; T CD4⁺ like Th1 and Th17 that became auto-reactive in the peripheral sites like lymph nodes, leading to demyelination by infiltrating the CNS and also secreting their specific pro-inflammatory cytokines^{11,12}; T CD8⁺ have a much higher number at the lesion sites because MHC class I molecules, which present antigens to these cells, is expressed in all nucleated cells; since Treg cells have a role in tolerance, when they are disrupted, immune cells become reactive¹⁰; B cells have a varied role in the progression of MS; they can activate T cells by processing and presenting antigens and also producing antibodies by plasma cells,^{5,11} which one way of treating MS is anti-CD20 treatment strategies (rituximab) that cause the depletion of B cells and a small population of circulating Th1 and Th17¹³ (Figure 1).

Innate immune cells play a significant role in the pathogenesis of MS; for instance: monocytes and macrophages secrete pro-inflammatory cytokines, DCs present antigens to T cells and activate them, whereas NKT cells

shift the immunological balance toward Th2 cells that can help in healing the disease.^{10,11}

2 | BERBERINE AND ITS BIOLOGICAL EFFECTS

In recent years, tremendous attention has been given to natural products due to their specific pharmacological activities for clinical applications in different national healthcare settings.¹⁴ Of these various compounds, Berberine with a molecular formula of C₂₀H₁₈NO₄ and a molar mass of 336.36 g/mol has emerged as a compound with extensive biological properties.¹⁴ Berberine is an isoquinoline alkaloid derivative found in the roots, rhizomes, and stem bark of plants within a variety of genera, including Annonaceae, Berberidaceae, Menispermaceae, Papaveraceae, Ranunculaceae, Rutaceae, and many others.¹⁵ Interestingly, Berberine appeared to be almost safe in usual doses, with a relatively low incidence of side effects.^{16,17} Berberine is a yellow odorless crystalline powder with an extremely bitter taste that is slightly soluble in ethanol or methanol, and sparingly soluble in water.¹⁶

Berberine, a traditional Chinese medicine, has long been used as an effective drug in treating metabolic disorders, and gastrointestinal, and endocrine diseases such as obesity, hypertension, fatty liver, type-2 diabetes, hyperlipidemia, diarrhea, and gout in humans and animals.^{16,18} Besides, accumulating evidence during the last two decades has addressed the polytrophic pharmacological potential of Berberine including antimicrobial, antioxidant, antitumor, anti-cardiovascular, antidiabetic properties, anti-hypertensive, immunosuppressive, anti-inflammatory, and neuroprotective activities.^{19,20}

Notably, the potential of Berberine in controlling different autoimmune disorders has been assessed in several experimental studies and clinical trials.²¹ As an anti-inflammatory and immunosuppressive, Berberine efficacy inhibits the inflammatory responses involved in clinically apparent autoimmune diseases MS, rheumatoid arthritis, psoriasis, and inflammatory bowel disease in human or animal models.²¹

The related mechanisms of Berberine against autoimmune diseases and associated inflammatory disorders mainly involve inhibiting inflammation, modulation of Th17/Treg balance favor to Treg, and modulation of Th1/Th2 balance favor to Th2.^{21–23} Of note, this effect is accompanied by inhibiting pro-inflammatory mediators and upregulating anti-inflammatory mediators^{20–22} (Figure 2).

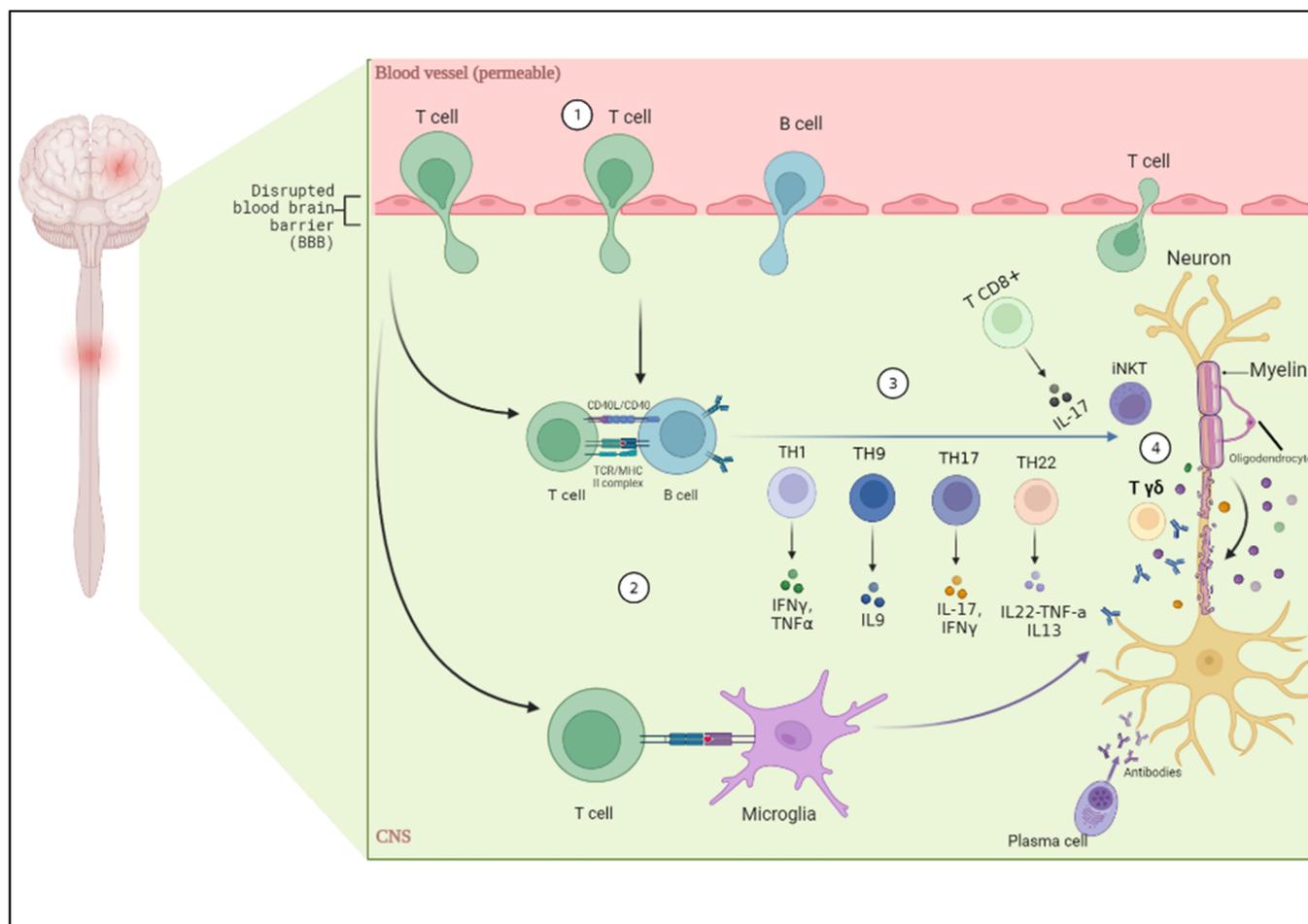


FIGURE 1 Immunopathology of multiple sclerosis: (1) blood–brain barrier (BBB) became permeable to lymphocytes. (2) T cell interacted with B cells and microglia to activate them. (3) Antibodies and cytokines released; antibodies released from plasma cells and cytokines were released from T cells. Th1 mainly released IFN- γ and TNF- α , Th9 released IL-9, Th17 released IL-17 and IFN- γ , Th22 released IL-22, TNF- α and IL-13 and T CD8⁺ released IL-17. (4) With these cytokines and cells demyelination of neurons occurred.

3 | ROLE OF DCs IN THE PATHOGENESIS OF MS AND MODULATORY EFFECTS OF BERBERINE ON DCs

Antigen-presenting cells (APCs) process and present antigens on MHC molecules for recognition by T cells via the TCR. DCs as a special subclass of APCs are important for the initiation and regulation of adaptive immunity, as well as in the triggering of autoimmunity.^{24,25} Human DCs have six subsets including blood versus tissue DCs, plasmacytoid DCs (pDCs), classical DCs (cDCs), CD1a⁺ CD14⁺ tissue DCs, Langerhans cells, and monocyte-derived DCs.²⁶

DCs have crucial roles in the development of the T cell repertoire as well as the activation and polarization of myelin-specific T cells in the peripheral and CNS in the context of MS and its model EAE.²⁷ MS is a complex autoimmune disease in which leukocytes, especially DCs

infiltrate into the CNS of patients, causing demyelination and axonal damage as a result.²⁸

Anatomic and flow cytometry studies relying on surface markers characteristic of peripheral DCs, have identified the presence of DCs in the CNS as indisputable.^{29,30} There are differences in the increasing presence and activation of DCs in the CNS of MS patients. Studies showed that the average number of DCs within the CSF was increased but it was not related to the severity of the disease as well as more DCs were seen in patients in the early phases of optic neuritis. In addition, CD11b⁺ DCs in CSF became more activated with increasing levels of costimulatory molecules and MHC class II.^{31,32}

There are some abnormal characteristics of circulating DC precursors and DCs from the blood of MS patients such as producing more TNF- α , IL-6, and IFN- γ . Moreover, the blood DCs of MS patients showed a pro-inflammatory profile including more IL-12 and TNF- α

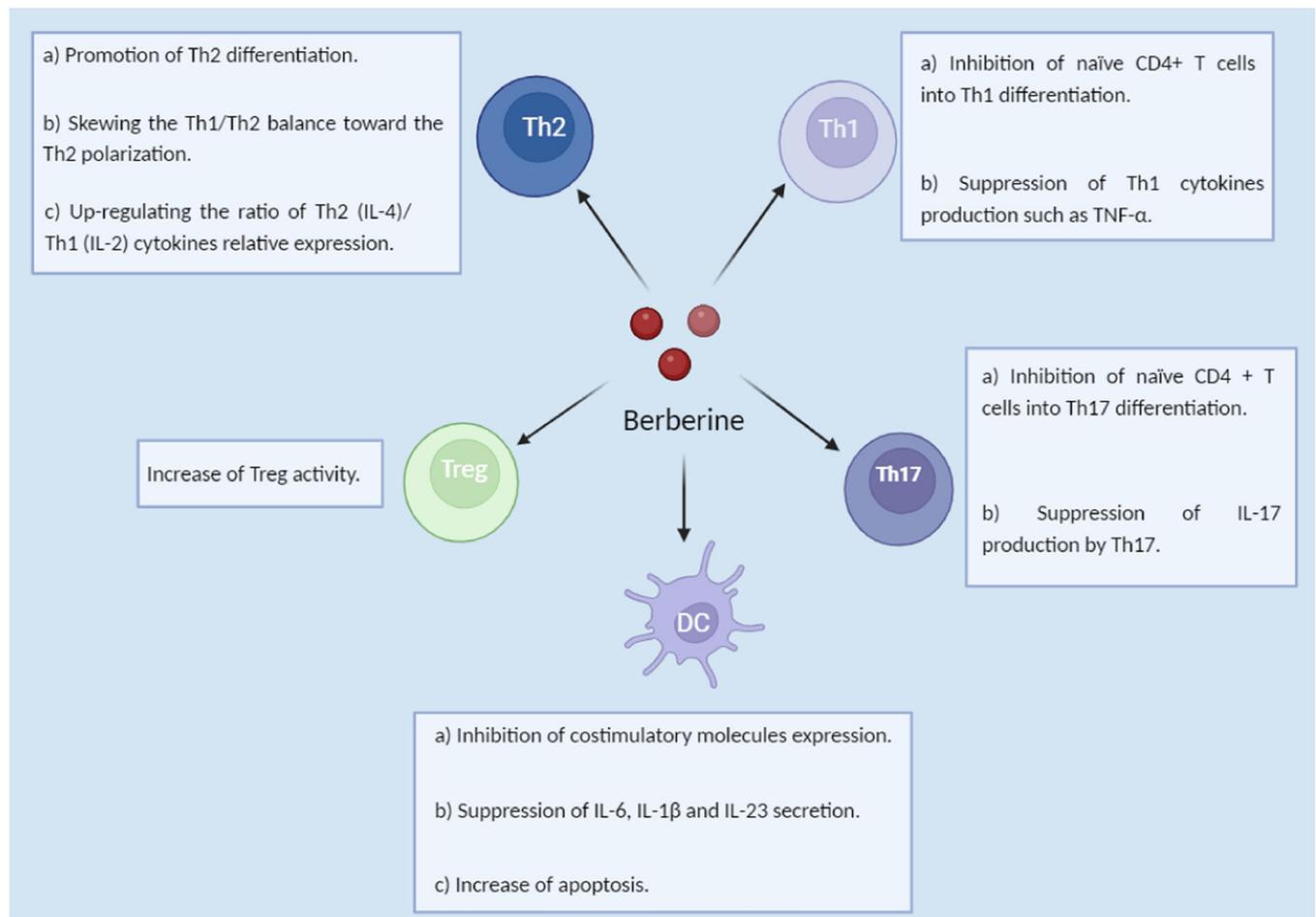


FIGURE 2 The overall effects of Berberine on immune cells: Berberine can regulate immune cells in different ways to moderate multiple sclerosis.

production along with costimulatory molecule upregulation.^{33,34} Furthermore, differences in DC function were found such as the capability of polarizing CD4⁺T cells toward IFN- γ production in a mixed lymphocyte reaction and an elevation of IL-23 secretion from monocyte-derived DCs suggests supporting abnormal Th17 polarization of T cells.^{35,36} Another study showed that a reduction of blood CD11b⁺DCs was found in secondary progressive MS (SPMS) compared to relapsing-remitting MS (RRMS)³⁷ patients (Figure 3).

Experimental evidence demonstrated the importance of DCs as an initiator of auto-reactive T-cell responses.³⁸ The role of DCs in MS pathogenesis is complicated because DCs are heterogeneous with a range of functional phenotypes.³⁹ Plasmacytoid DCs have two different subsets including type 1 (pDC1) with high levels of CD123, low levels of CD86 and TLR2 which are the source of IFN- α and induce IL-10 producing T cells, and type 2 (pDC2) with low levels of CD123, high levels of CD86 and TLR2 that, secret IL-6 and TNF- α and direct naïve T cells toward IL-17 secreting Th17 cells. The

pDC1/pDC2 ratio in MS patients is a noteworthy shift to pDC2.⁴⁰ Moreover, some mechanisms, related to DCs, such as type 1 IFN production by pDCs, might lead to pathogenesis in autoimmune disease.⁴¹ Furthermore, increases in the expression of the CCR5 in cDCs of CSF have been observed.^{32,42}

Abnormalities of DCs contribute to the pathology and response to treatment of MS. Considering the role of DCs in the pathogenesis of MS; they can be used as a therapeutic target in MS. Actually, the current immunomodulatory therapies used for the treatment of MS affect DC function.⁴³ For example, PDL1, a molecule causes the inhibition of DCs signaling to T cells.⁴⁴ Currently, Glatiramer acetate is used as an immunomodulatory drug to treat MS, which is associated decreasing in TNF- α and IL-12p70 production by monocyte-derived DCs and reduced CD40 expression and pro-inflammatory activity in pDCs.⁴⁵

Mature DCs were found after LPS stimulation more sensitive to berberine than immature DCs. So, the expression of CD80/CD86 and IL-12 was low in the

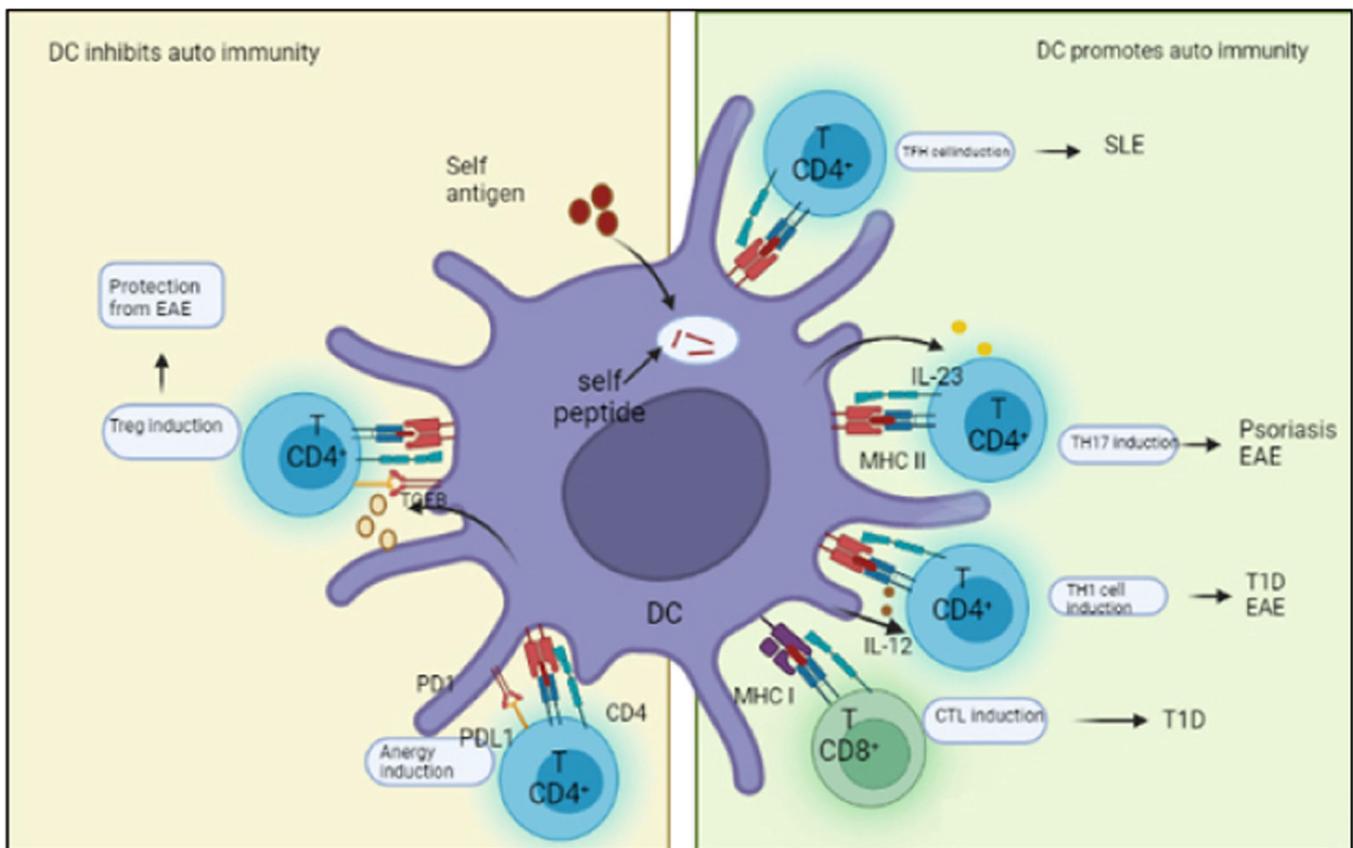


FIGURE 3 Roles of DCs in autoreactive T cell responses. Presentation of self-antigens to T cells in the context of programmed cells, the interaction between PD1 and PDL1 and TGF- β signaling cause Anergy of autoreactive T cells can promote their development into Treg. In contrast, presentation of self-antigens to T cells in the context of pro-inflammatory mediators such as IL-6, IL-12, and IL-23 promotes the development of self-reactive effector CD4⁺ T cells and CTLs. These self-reactive T cells might cause pathological autoimmune diseases, including EAE in mice, or systemic lupus erythematosus (SLE), psoriasis, and Type 1 diabetes (T1D) in patients. DC, dendritic cell.

presence of berberine in response to LPS stimulation.⁴⁶ Some effects of berberine on mature DCs stimulated with LPS were shown as inhibition of expression of costimulatory molecules including CD80, CD86, and CD40. Furthermore, the production of IL-23 was induced; in contrast, the levels of IL-6 and IL-1 β were decreased.⁴⁷ Also, berberine can activate DC apoptosis.⁴⁸ It is necessary to mention that more studies should be done to investigate the effect of berberine on DC function and related mechanisms.

4 | ROLE OF TH1 CELLS IN THE PATHOGENESIS OF MS

A subset of TCD4⁺ called Th1 cells is produced as a result of IL-12 and IFN- γ signals acting on naïve T cells.^{49,50} Th1 is often identified by the expression of surface markers including CXC chemokine receptor type 3 (CXCR3) and IL-12 receptor (IL-12R), as well as the production of a transcription factor called T-bet that is induced by

IL-12 and IFN- γ signaling.^{51–53} Th1 can produce variant cytokines such as IFN- γ , IL-2, TNF- α , and GM-CSF.^{54,55} IFN- γ , the main cytokine generated by Th1 cells, causes macrophages to become activated and fight intracellular pathogens.⁴⁹

Additionally, Th1 is primarily responsible for the development of autoimmune disorders like MS.⁵⁶ The autoreactive Th1 cells can penetrate CNS endothelium and run away from the regulation mechanisms of the immune system and directly or indirectly destroy the oligodendrocytes, myelin sheaths, and axons. Indirect damage is caused by the production of pro-inflammatory cytokines such as IFN- γ and IL-2⁵⁷ (Figure 4).

IFN- γ , as a signature cytokine produced by Th1 cells, is an important factor in the progression of MS.^{58,59} All CNS cells have IFN- γ receptors, which means they could be able to react to this cytokine.⁶⁰ Some studies have revealed that MS patients exhibit elevated amounts of TNF- α and IFN- γ generated by the Th1 cell type.⁶¹ Furthermore, other studies showed that MS patients with active lesions had higher IFN- γ values.⁶² MS risk may

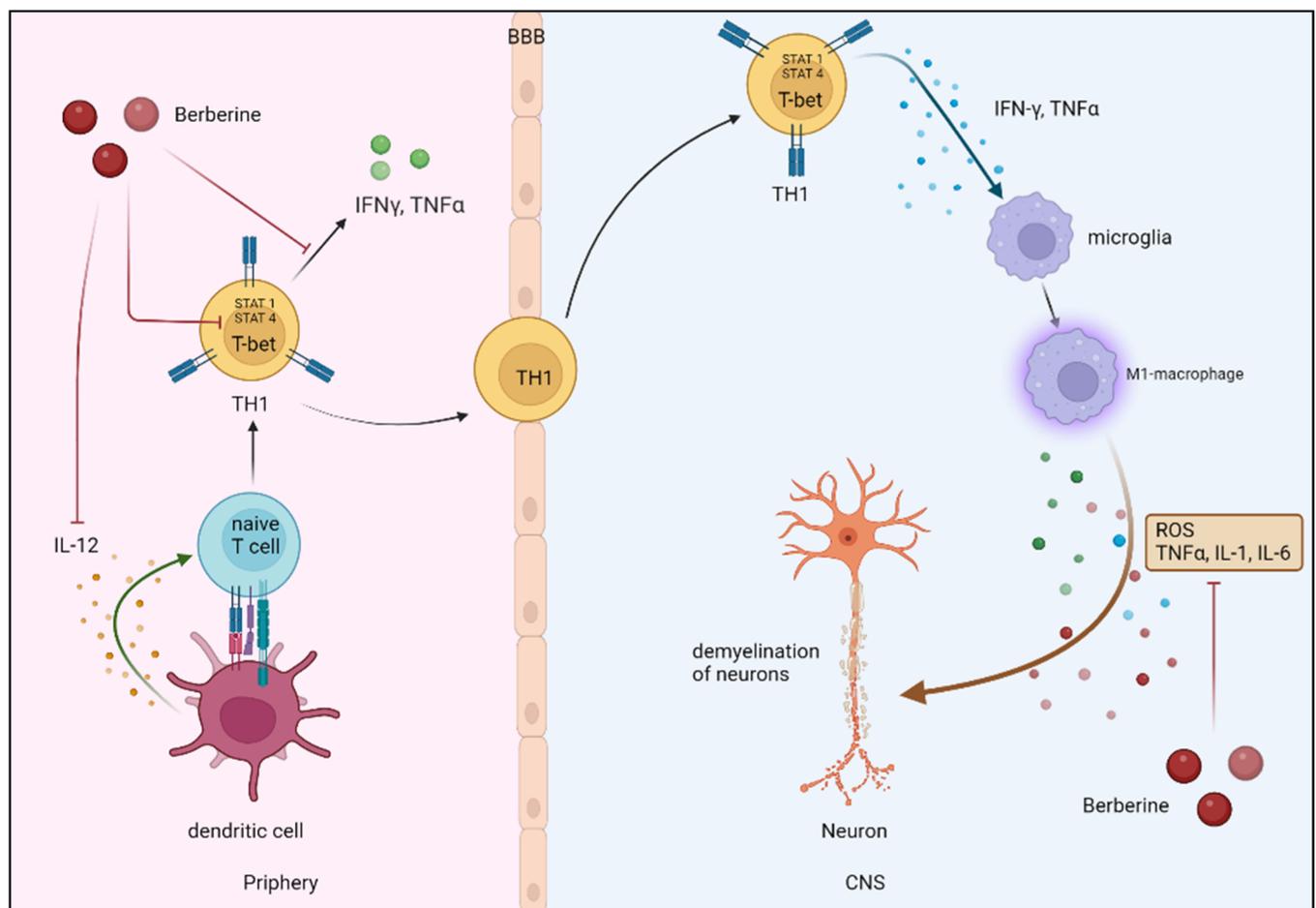


FIGURE 4 Naïve T cells differentiate into Th1 by presenting the antigen and producing cytokines like IL-12 from dendritic cells. Th1 cells can cross the blood–brain barrier (BBB) and escape from immune system regulatory mechanisms. As a result, it activates CNS-resident macrophages (microglia) by generation of cytokines like IFN- γ and changes them into M1-phenotype cells that can cause demyelination of neurons by producing ROS and inflammatory cytokines. Berberine can reduce the impact of IL-12 on naïve T cells, which in turn decreases the expression of STAT 1, STAT 4, and T-bet and consequently causes the reduction in the Th1 populations. Additionally, berberine can limit the production of inflammatory cytokines from Th1 and M1-macrophages and also neutralize the effect of ROS produced by M1-macrophage. CNS, central nervous system; ROS, reactive oxygen species.

rise due to IFN- γ gene polymorphism.^{63,64} Th1 can induce the activation of CNS resident microglia, and cause the distinction of these cells to inflammatory phenotype (M1-like) through the production of IFN- γ .^{65,66} This activated macrophage can participate in the progress of the disease via the generation of inflammatory cytokines and reactive oxygen species (ROS).⁶⁷

Inconsistently displayed that Th1 may act as a protective agent in the pathogenesis of MS. A study done on EAE animal models, observed that animals with IFN- γ defects suffered from a more severe form of the disease compared to animals with normal amounts of IFN- γ .^{68,69} Moreover, Ni et al, find out that IFN- γ leads to the sustainability of the blood–brain barrier (BBB).⁷⁰

Nicotinamide adenine dinucleotide (NAD) levels are influenced by Th1-generated cytokines such IFN- γ , IL-1, and TNF- α through induced activation of indoleamine

2,3-dioxygenase (IDO) in the APCs such microglia, macrophage, and DCs during chronic CNS inflammation.⁷¹ IDO discharges the tryptophan, a substrate for NAD synthesis, and causes the reduction of its extracellular concentration.⁷¹ A shortage of extracellular tryptophan causes decreasing in the proliferation of autoreactive T cells.^{72,73} However, contradictory abnormal excessive IDO activity might miss neuronal CNS cells famished for external NAD sources. Therefore, demyelination recovery in MS depends on maintaining a homeostatic balance of IDO activity.⁷¹

Berberine can prevent the differentiation of naïve CD4⁺ T cells into Th1 via decreased phosphorylation of STAT1 and STAT4 that activated following IL-12 signaling, as well as suppression of T-bet representation.^{23,74–76} It was demonstrated in EAE mice that berberine therapy resulted in a decrease in the number

of Th1 cells by blocking the effects of IL-12 signaling on naive T cells.⁷⁵ The animal model of autoimmune peripheral demyelinating diseases like Guillain–Barre syndrome, known as experimental autoimmune neuritis (EAN), also revealed that berberine can suppress the production of Th1 cytokines such as TNF- α and result in a reduction in the intensity of the sickness.⁷⁷ Inversely, it was found that berberine may cause Th1 differentiation by inducing the production of IL-12.⁷⁸ It is thought that these actions of berberine lead to the treatment of disorders like asthma, in which Th2 plays a prominent role.⁷⁹

5 | ROLE OF TH17 CELLS IN THE PATHOGENESIS OF MS

Th17 cells confer powerful protective immunity to infectious organisms; they also have been found to play crucial pathogenic roles in the inflammatory response and demyelinating lesions within the CNS of MS patients.⁸⁰ Th17 cells are identified by the production of their hallmark cytokine IL-17A and the expression of the transcription factor ROR- γ t.⁸¹ In addition to IL-17, Th17 cells produce other inflammatory cytokines such as IL-21 and IL-22.⁸²

TGF- β , in combination with pro-inflammatory and pleiotropic cytokines such as IL-6 and IL-23, is required to induce Th17 cell lineage commitment via expression of transcription factor ROR- γ t.^{83,84} The pathogenic role of Th17 cells in autoimmune diseases has emerged from studies that indicate that IL-17-deficient mice are lowly susceptible to EAE, and treatment with IL-17R antagonist or IL-17 neutralizing antibody led to the amelioration of EAE.²¹ Additional supporting data for the Th17 came from the study showing that Th17-axis cell phenotypes and IL-17 concentration are increased in MS patients with active disease in comparison to healthy subjects.^{82,85} Further evidence demonstrated that augmented expression of IL-17 was observed in patients in both brain lesions and mononuclear cells isolated from blood and cerebrospinal fluids.⁸⁶ More recently, an increase in the production of IL-17 is also observed in lymphocytes derived from mice with EAE.^{86,87} Consistent with these observations, IL-17 transcript is upregulated in MS lesion⁶² and MS activity is correlated with an increased number of Th17 cells in the patient's blood.⁸⁸ The pathogenic effect of Th17 in the EAE model is further confirmed by findings that deficiency or neutralization of IL-17A delays the onset and reduces the incidence and severity of relapses.⁸⁶

Other proposed pathological functions of IL-22 and IL-17, two of the cytokines produced by Th17 cells, include the disruption of the blood-brain barrier *in vitro* and *in vivo*

and the killing of human neurons.^{82,89} These results are in line with our recent data showing effector molecules secreted by Th17 cells induce a strong pro-inflammatory response and suppress an anti-inflammatory response in astrocytes⁹⁰ and have been associated with disease onset, progression, and relapse.⁹¹

Th17 responses are critically involved in the pathogenesis of several autoimmune diseases, so reviewing the effects of Berberine on these cells can be valuable for the treatment of these diseases. An experiment conducted in EAE suggested Berberine can inhibit the differentiation and function of Th17 cells through direct actions on the JAK/STAT signaling pathway; in this way, Berberine downregulates STAT3 phosphorylation and ROR- γ t expression, during Th17 cells differentiation. In addition, Berberine indirectly influences Th17 cells by inhibition of NF- κ B activity in CD11b⁺ APCs, which correlates with the downregulation of costimulatory molecules and suppression of cytokine IL-6 production; therefore, Berberine can improve EAE.⁷⁵ Another study confirmed that Berberine directly suppressed IL-17 production by Th17 cells and differentiation of Th17 cells via its indirect effect on DCs from Vogt-Koyanagi-Harada patients.⁴⁷ In type 1 diabetes, the Berberine treatment of NOD mice prevented Th17 differentiation by increasing ERK1/2 activity, which implies that ERK may have a negative regulatory role in Th17 differentiation. ERK inhibits Th17 Differentiation by downregulation of STAT3 and ROR- γ t signaling pathway.⁹² Research directed by Mengfan Yue et al. has shown that oral Berberine improves collagen-induced arthritis (CIA) in rats, an animal model for human rheumatoid arthritis, by suppressing Th17 cell responses, which was correlated with the stimulation of cortistatin (an immunoregulatory neuropeptide) generation from the gut. Intestinal mRNA and protein levels of cortistatin enhanced in Berberine-treated rats and it negatively attenuated the systemic responses of Th17 cells.⁹³ Further evidence also characterized that Berberine could decline excessive responses of Th17 cells and the levels of Th17 cytokines in a rat model of experimental autoimmune myocarditis (EAM), so EAM ameliorated by BRR. The effect of Berberine on Th17 cells is mediated by the downregulation of STAT3 phosphorylation.⁷⁴ Another study on mice with EAE indicated that Berberine suppresses the differentiation of naive CD4⁺ T cells into pathogenic Th17 cells through direct actions on the JAK/STAT pathway, resulting in the amelioration of inflammation in EAE-induced mice.⁷⁵ We are confirmed by a study on EAE mice that showed treatment with Berberine significantly downregulates the expression of transcription factors (ROR γ t) and pro-inflammatory cytokine derived from Th17 such as IL-17, which was accompanied by reduced CNS inflammation

and demyelination.²³ Collectively, results from studies point to the promising role Berberine has in suppressing pathogenic Th17 cells in EAE.

6 | ROLE OF TH2 CELLS IN THE PATHOGENESIS OF MS

Naive T cells can be induced to differentiate Th2, a distinct subset of Th cells characterized by expression of the transcription factor (GATA-3), which is associated with humoral immunity and allergic immune response.⁹⁴ Effector Th2 cells secrete a profile of potent anti-inflammatory cytokines, including IL-4, IL-5, IL-10, and IL-13, and have been implicated in the regulation of autoimmune diseases.⁹⁴ Th2-associated cytokines tend to mediate anti-inflammatory humoral response and immune suppression via the inhibition of Th1 cytokine production.⁹⁵ Recent studies have shown that among the T effector subsets not only Treg, but also Th2 cells contribute to recovery from disease by controlling the expansion and activation of autoreactive CD4⁺ T effector cells.⁹⁶ It is noteworthy that the Th1 and Th2 subsets mutually antagonize one another's function and breaking the Th1/Th2 balance with a predominance of Th1 was considered to be important in the initiation and perpetuation of autoimmunity of MS. Additionally, a shift from a Th1 towards a Th2 cytokine profile have been associated with inflammation reduction and improvement of autoimmune conditions.^{97,98}

The protective role of Th2 cytokines against inflammatory disease comes from studies that indicate enhancement of a Th2 cytokine will ameliorate or prevent inflammatory disorders, whereas decreased production of Th2 cytokines may be involved in the pathogenesis of several inflammatory disorders.^{99,100} In addition, the adoptive transfer of myelin proteolipid protein (PLP)-specific Th2 cells has been demonstrated to inhibit ongoing EAE and as well as abrogate established disease.^{101,102} Furthermore, recovery from EAE is associated with an increase in the expression of IL-4 and Th2 cell production in the CNS, further supporting the hypothesis that Th2 cells can prevent the propagation of inflammation in EAE/MS.^{101–103} The mechanisms by which Th2 cells suppress EAE control the development and activity of encephalitogenic Th1 cells and the induction of the alternative (M2) type of macrophages/microglia.^{94,104}

Berberine could promote the differentiation of Th2 cells which play a protective role in disease development and alleviating autoreactive inflammatory responses.^{21,23} The potential for Berberine to favor Th2 type response may be an effective therapeutic agent to ameliorate

Th1-skewed inflammatory and autoimmune diseases, such as MS, RA, Crohn's disease, and type 1 diabetes.¹⁰⁵

An *in vitro* study demonstrated that Berberine exhibits its immunomodulatory and anti-inflammatory potential by skewing the Th1/Th2 balance toward the Th2 polarization in mouse primary splenocytes.¹⁰⁵ This anti-inflammatory effect is mediated by upregulating the ratio of Th2 (IL-4)/Th1 (IL-2) cytokines relative expression.¹⁰⁵ Investigation of Berberine administration on modulating the immune response in the experimental model of MS showed that the expression of transcription factors and Th2 and Treg cytokines such as IL-4, STAT6, GATA3, IL-10 in splenocytes and lymph nodes from EAE mice is significantly increased in Berberine-treated groups.²³ Shifting the balance of Th1/Th2 towards Th2 is associated with reduced CNS inflammation and demyelination in treated groups.²³ Therefore, Berberine could be helpful in treating autoimmune and inflammatory diseases by shifting the Th1/Th2 balance toward Th2 polarization.

7 | ROLE OF TREG CELLS IN THE PATHOGENESIS OF MS

Treg cells have anti-inflammatory effects and maintain tolerance to self-antigens.⁵⁵ In terms of immunology, MS is associated with the dysfunction of Treg and increases the response of Th1 and Th17 cells.¹⁰⁶ Research has determined that the regulatory activity of CD4⁺ CD25^{hi} T-cell is impaired in MS patients.¹⁰⁷ FoxP3 is a specific transcription factor of Treg cells that plays a crucial role in the development and function of these cells.¹⁰⁸ The brain biopsy of patients with MS has shown that 30% of the lesions do not express FoxP3. Fas a cell surface receptor for apoptosis, has been shown that increased Tregs in MS brain biopsies indicate enhancement of the susceptibility to apoptosis.¹⁰⁹ Disruption of Treg cell function during the development of the disease can be modulated by CD28 and CTLA-4. The research in EAE has identified that expression of CD28, colocalizing with T cell receptor subunit CD3, increases during relapse.¹¹⁰ Use of IDO metabolite, which leads to an increase in Treg cell number, leads to considerable improvement of MOG-induced EAE.¹¹¹

In relapsing-remitting EAE models, Treg cell depletion can increase the severity of the acute phase of the disease and prevent its recovery.^{112,113} Some MS-related studies have indicated that although the number of Treg cells is normal, they have functional deficiencies, and in patients with MS, Treg cells have less power to suppress IL-17 production compared to healthy subjects. Other studies show a two- to threefold reduction in the number

of Treg cells in the MS exacerbation phase and an increase in the number of these cells in the regression phase, and there is an inverse relationship between the severity and duration of the disease with the number of Treg cells.^{114,115} Regulatory activity of the Treg-producing IL-10 cells is characterized by the transfer of IL-10-producing Treg cells to EAE mice, and treatment with IL-10 antagonists also increases the severity of EAE.¹¹⁶

The results of a recent study suggest that Treg cells with the expression of the transcription factor Forkhead box protein A1 (FOXA1) play a role in controlling the immune response. FOXA1 is responsible for the expression of Programmed cell death ligand 1 (PD-L1), which mediates the destruction of T-cells activated by binding to the PD-1R receptor; therefore, the active transfer of Treg FOXA1 cells inhibits EAE.¹¹⁷ Treg cells protect neurons by direct induction of apoptosis in the inflammatory microglial cells of M1, as well as the change in the phenotype of these cells towards M2 macrophage.¹¹⁸

Another study has shown that Treg cells enhance the expression of astrocyte-derived neurotrophic factors, the brain-derived neurotrophic factor, and the glial cell-derived neurotrophic factor, which promotes remyelination and brain repair. It also results in significant improvement in the function of the neural cells with their ability to inhibiting produce of ROS and glutamate.^{119,120}

In general, promoting the activity of Treg cells or using their cytokines to treat MS disease is worthy of attention.

Recent research has suggested the use of Berberine as a cure for some autoimmune diseases such as EAE,^{121,122} Type 1 diabetes,¹²³ and Rheumatoid arthritis.¹²⁴ The effect of Berberine on spleen tissue Treg cells has been studied by our lab. The results of this study showed that Berberine increases the activity of Treg cells and also induces the production of TGF- β and IL-10 by splenocytes.²³ As reported by Li et al., The differentiation of Th1 and Th17 cells was significantly suppressed by the addition of Berberine, while Berberine did not have a significant effect on Treg cell differentiation.¹²⁵

8 | ROLE OF OTHER IMMUNE CELLS IN THE PATHOGENESIS OF MS AND MODULATORY EFFECTS OF BERBERINE ON THESE CELLS

In recent years, it has been discovered that there is a distinct subset of IL-9-producing effector CD4⁺ T cells. Even though the fact that IL-9 was formerly connected to a Th2 response, recent research has redefined IL-9-producing CD4⁺ T cells as Th9 cells^{126,127} Comparing

mice with EAE receiving Th9 cells to mice receiving Th1 and Th17 cells, showed that mice receiving Th9 cells exhibited less lymphocyte infiltration in the meninges.¹²⁸ Additionally, the protective function of IL-9 was demonstrated in IL-9R knockout mice, which exhibited a severe form of EAE.¹²⁹ Also, Ruocco et al. discovered an inverse relationship between IL-9 levels in the CSF of RRMS patients and measures of inflammatory activity, neurodegeneration, and the development of MS-related disability.¹³⁰ However, some studies showed that IL-9 neutralization and IL-9R deficiency reduced EAE.^{131,132} It has been observed that adoptively transfer of PLP₁₈₀₋₁₉₉ peptide-specific T cells from wild-type mice or immunization with the PLP₁₈₀₋₁₉₉ peptide resulted in significantly less severe EAE and lower levels of IL-17 and IFN- γ in IL-9 knockout mice compared to wild-type mice.¹³³ However, the role of Th9 and IL-9 in the pathogenesis of MS is not clearly defined.

Th22 is characterized by the release of several cytokines, including the most significant one, IL-22, as well as IL-13 and TNF- α .¹³⁴ Beyeen et al showed a connection between the IL-22R α 2 gene and an increased chance of MS.¹³⁵ Additionally, patients with MS and neuromyelitis Optica (NMO) showed an increase in IL-22 and Th22 cells.¹³⁶ Kebir et al showed that the IL-22R was overexpressed in the brains of MS patients. They found that IL-22 and IL-17A worked together to damage the integrity of BBB tight junctions by lowering the expression of occludin in endothelial cells.⁸⁹ More recent research revealed that relapsing MS patients had higher serum levels of IL-22 than healthy donors,¹³⁶⁻¹³⁹ while during the recovery phase of acute EAE, IL-22 levels decreased.¹⁴⁰ However, another study showed that mice lacking IL-22 are completely susceptible to EAE induced by the MOG₃₅₋₅₅ peptide.¹⁴¹ Therefore, more research is required to determine the exact role of IL-22 in autoimmune inflammatory disorders of the CNS.

Although much focus on MS pathology has centered on CD4 T cells, data suggest that CD8 T cells play a role in MS. Studies have suggested that the MBP-specific CD8⁺ T lymphocytes may increase inflammation in the brain.^{142,143} The primary lymphocytes found in the CNS brain lesions of MS patients and mice with EAE were found to be CD8⁺ T cells.¹⁴² EAE in mice might be induced by the adoptive transfer of CD8⁺ enriched MOG-specific T cells.⁵⁴ In another investigation, it was discovered that people with MS had more circulating CD8⁺ CD20⁺ T memory cells that were specific to myelin antigens than control subjects. These T cells may be very pathogenic since CD20 expression on them is connected with the upregulation of activation markers, pro-inflammatory cytokines, and adhesion molecules. Adoptive transfer of CD20⁺ T cells in EAE mice, which led to

the degradation of brain tissue integrity and exacerbation of disease severity, supported this concept. Following anti-CD20 treatment, the fraction of memory and CD20⁺CD8⁺ T cells that are specific for myelin antigens was considerably decreased, showing that the deletion of CD20⁺ T cells has therapeutic promise for MS.¹⁴⁴ Cytokine-expressing CD8 T cells are also related to MS. In contrast to inactive lesions, where only a small number of CD8 T cells were IL-17 positive, active MS lesions had an increase in these cells in the perivascular regions.¹⁴⁵ All of these CD8 T cells that secrete IL-17 are members of the CD161^{high} subset of CD8 cells.¹⁴⁶

NKT cells, also known as natural killer T cells, are a heterogeneous group of T cells that express NK cell surface antigens. Type I NKT, also known as invariant NKT or iNKT, type II (nonclassical) NKT, and NKT-like cells are the three main populations of NKT cells.¹⁴⁷ iNKT cells have an invariant V α 24J α 18 T-cell receptor and identify self- and foreign-derived lipids provided by CD1d as cognate antigens.^{148,149} Changing the iNKT cell population in the peripheral blood of MS patients does not follow a certain pattern in different forms of MS or during different treatments of the disease,^{150–152} but there is probably a frequency change among different iNKT cell subsets.¹⁵³ Depending on the form of MS, cytokine production in subtypes of iNKT cells is different.^{150,153,154} iNKT cells may perhaps be a mediator of the immune regulatory impact of IFN- β therapy in MS.¹⁵⁵ It was reported in several studies that iNKT cells can protect mice from EAE by modulating CD4⁺ Th1- and Th17-mediated immune responses, but with different mechanisms.^{156–158} Besides, some studies show that iNKT cells are more protective against the myelin-specific Th1 responses after stimulation with alpha-galactosylceramide (α -GalCer).^{159–161} Also, another research demonstrates that after receiving α -GalCer, iNKT cells expressing FoxP3 showed protective effects in EAE.¹⁶²

$\gamma\delta$ T cells are a discrete group of lymphocytes that express TCR, which is made up of two glycoprotein chains on the surface known as the γ and δ TCR chains. Several studies have reported that $\gamma\delta$ T cells present in MS plaques of MS patients (with enrichment to a TCR-restricted repertoire)¹⁶³ and an increase in $\gamma\delta$ T lymphocytes in the CSF, which is associated with their rise in peripheral blood.¹⁶⁴ $\gamma\delta$ T cells isolated from the CNS can be expanded, but only in individuals with the recently developed disease and not in those with chronic MS; whereas expansion of CD16⁺ cytotoxic $\gamma\delta$ T cells in MS patients was mainly seen during the progressive phase of the disease.¹⁶⁵ $\gamma\delta$ T cells may play a part in demyelinating diseases; however, this is unclear.

Whether Berberine can affect these cells has not yet been discovered. Hence more investigations need to understand how berberine can affect these cells to change the MS state.

AUTHOR CONTRIBUTIONS

Esmail Yazdanpanah, Sepehr Dadfar, Alireza Shadab, Niloufar Orooji, MohammadHossein Nemati, Alireza Pazoki, and Dariush Haghmorad wrote the main manuscript. Sepehr Dadfar, Niloufar Orooji, and MohammadHossein Nemati designed the figure. Seyed-Alireza Esmaili, Rasoul Baharlou, and Dariush Haghmorad reviewed the manuscript and figure.

CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

ORCID

Dariush Haghmorad  <http://orcid.org/0000-0002-9876-4943>

REFERENCES

- Dendrou CA, Fugger L, Friese MA. Immunopathology of multiple sclerosis. *Nat Rev Immunol.* 2015;15(9):545-558. doi:10.1038/nri3871
- Howard J, Trevick S, Younger DS. Epidemiology of multiple sclerosis. *Neurol Clin.* 2016;34(4):919-939. doi:10.1016/j.ncl.2016.06.016
- Azami M, YektaKooshali MH, Shohani M, Khorshidi A, Mahmudi L. Correction: epidemiology of multiple sclerosis in Iran: a systematic review and meta-analysis. *PLoS One.* 2019;14(7):e0219466. doi:10.1371/journal.pone.0219466
- Van Liew C, Dibble LE, Hunt GR, Foreman KB, Peterson DS. Protective stepping in multiple sclerosis: impacts of a single session of in-place perturbation practice. *Mult Scler Relat Disord.* 2019;30:17-24. doi:10.1016/j.msard.2019.01.054
- Dobson R, Giovannoni G. Multiple sclerosis—a review. *Eur J Neurol.* 2019;26(1):27-40. doi:10.1111/ene.13819
- Farnoush R, Aliloo L, Sahebolzamani M, Rahmani A. Educational, psycho mental and socio economical needs of an Iranian cohort with multiple sclerosis. *Oman Med J.* 2010;25(1):22-25. doi:10.5001/omj.2010.6
- Kobelt G, Thompson A, Berg J, Gannedahl M, Eriksson J. New insights into the burden and costs of multiple sclerosis in Europe. *Mult Scler J.* 2017;23(8):1123-1136. doi:10.1177/1352458517694432
- Lassmann H, Brück W, Lucchinetti CF. The immunopathology of multiple sclerosis: an overview. *Brain Pathol.* 2007;17(2):210-218. doi:10.1111/j.1750-3639.2007.00064.x
- Mahmoudi MB, Mahmoudi M, Rab SZT, et al. Calcium intervention ameliorates experimental model of multiple sclerosis. *Oman Med J.* 2014;29(3):185-189. doi:10.5001/omj.2014.46
- Dargahi N, Katsara M, Tselios T, et al. Multiple sclerosis: immunopathology and treatment update. *Brain Sci.* 2017; 7(7):78. doi:10.3390/brainsci7070078

11. Hemmer B, Kerschensteiner M, Korn T. Role of the innate and adaptive immune responses in the course of multiple sclerosis. *Lancet Neurol.* 2015;14(4):406-419. doi:10.1016/S1474-4422(14)70305-9
12. Haghmorad D, Yousefi B, Eslami M, et al. Oral administration of myelin oligodendrocyte glycoprotein attenuates experimental autoimmune encephalomyelitis through induction of Th2/Treg cells and suppression of Th1/Th17 immune responses. *Curr Issues Mol Biol.* 2022;44(11):5728-5740. doi:10.3390/cimb44110388
13. Bar-Or A, Li R. Cellular immunology of relapsing multiple sclerosis: interactions, checks, and balances. *Lancet Neurol.* 2021;20(6):470-483. doi:10.1016/S1474-4422(21)00063-6
14. Majdalawieh AF, Yousef SM, Abu-Yousef IA, Nasrallah GK. Immunomodulatory and anti-inflammatory effects of berberine in lung tissue and its potential application in prophylaxis and treatment of COVID-19. *Front Biosci (Landmark Ed).* 2022;27(5):166. doi:10.31083/j.fbl2705166
15. Wang H, Zhang H, Gao Z, Zhang Q, Gu C. The mechanism of berberine alleviating metabolic disorder based on gut microbiome. *Front Cell Infect Microbiol.* 2022;12:854885. doi:10.3389/fcimb.2022.854885
16. Lei W, Sagada G, Wang C, et al. Berberine in fish nutrition: impact on hepatoenteric health, antioxidative and immune status. *Front Mar Sci.* 2022;9:1-16. doi:10.3389/fmars.2022.967748
17. Huang DN, Wu FF, Zhang AH, Sun H, Wang XJ. Efficacy of berberine in treatment of rheumatoid arthritis: from multiple targets to therapeutic potential. *Pharmacol Res.* 2021;169:105667. doi:10.1016/j.phrs.2021.105667
18. Zhang L, Wu X, Yang R, et al. Effects of berberine on the gastrointestinal microbiota. *Front Cell Infect Microbiol.* 2020;10:588517. doi:10.3389/fcimb.2020.588517
19. Zhong XD, Chen LJ, Xu XY, et al. Berberine as a potential agent for breast cancer therapy. *Front Oncol.* 2022;12:993775. doi:10.3389/fonc.2022.993775
20. Mohammadian Haftcheshmeh S, Momtazi-Borojeni AA. Berberine as a promising natural compound for the treatment of periodontal disease: a focus on anti-inflammatory properties. *J Cell Mol Med.* 2021;25(24):11333-11337. doi:10.1111/jcmm.17019
21. Ehteshamfar SM, Akhbari M, Afshari JT, Seyedi M, Nikfar B. Anti-inflammatory and immune-modulatory impacts of berberine on activation of autoreactive T cells in autoimmune inflammation. *J Cell Mol Med.* 2020;24(23):13573-13588. doi:10.1111/jcmm.16049
22. Shen P, Jiao Y, Miao L, Chen JH, Momtazi-Borojeni AA. Immunomodulatory effects of berberine on the inflamed joint reveal new therapeutic targets for rheumatoid arthritis management. *J Cell Mol Med.* 2020;24(21):12234-12245. doi:10.1111/jcmm.15803
23. Tavaf MJ, Soltanmohammadi A, Zargarani S, et al. Berberine promotes immunological outcomes and decreases neuroinflammation in the experimental model of multiple sclerosis through the expansion of Treg and Th2 cells. *Immun Inflamm Dis.* 2023;11(1):e766. doi:10.1002/iid3.766
24. Steinman RM. Decisions about dendritic cells: past, present, and future. *Annu Rev Immunol.* 2012;30:1-22.
25. Hardin JA. Dendritic cells: potential triggers of autoimmunity and targets for therapy. *Ann Rheum Dis.* 2005;64(Suppl 4):iv86-iv90.
26. Collin M, McGovern N, Haniffa M. Human dendritic cell subsets. *Immunology.* 2013;140(1):22-30.
27. Mildner A, Jung S. Development and function of dendritic cell subsets. *Immunity.* 2014;40(5):642-656.
28. Schmidt H, Williamson D, Ashley-Koch A. HLA-DR15 haplotype and multiple sclerosis: a HuGE review. *Am J Epidemiol.* 2007;165(10):1097-1109.
29. Nataf S, Strazielle N, Hatterer E, Mouchiroud G, Belin MF, Gherzi-Egea JF. Rat choroid plexuses contain myeloid progenitors capable of differentiation toward macrophage or dendritic cell phenotypes. *GLIA.* 2006;54(3):160-171.
30. Fischer H-G, Reichmann G. Brain dendritic cells and macrophages/microglia in central nervous system inflammation. *J Immunol.* 2001;166(4):2717-2726.
31. Kivisäkk P, Mahad DJ, Callahan MK, et al. Expression of CCR7 in multiple sclerosis: implications for CNS immunity. *Ann Neurol.* 2004;55(5):627-638.
32. Pashenkov M, Teleshova N, Kouwenhoven M, et al. Elevated expression of CCR5 by myeloid (CD11c+) blood dendritic cells in multiple sclerosis and acute optic neuritis. *Clin Exp Immunol.* 2002;127(3):519-526.
33. Huang Y-M, Xiao B-G, Özenci V, et al. Multiple sclerosis is associated with high levels of circulating dendritic cells secreting pro-inflammatory cytokines. *J Neuroimmunol.* 1999;99(1):82-90.
34. Karni A, Abraham M, Monsonego A, et al. Innate immunity in multiple sclerosis: myeloid dendritic cells in secondary progressive multiple sclerosis are activated and drive a proinflammatory immune response. *J Immunol.* 2006;177(6):4196-4202.
35. Vaknin-Dembinsky A, Balashov K, Weiner HL. IL-23 is increased in dendritic cells in multiple sclerosis and down-regulation of IL-23 by antisense oligos increases dendritic cell IL-10 production. *J Immunol.* 2006;176(12):7768-7774.
36. Stasiulek M, Bayas A, Kruse N, et al. Impaired maturation and altered regulatory function of plasmacytoid dendritic cells in multiple sclerosis. *Brain.* 2006;129(5):1293-1305.
37. López C, Comabella M, Al-Zayat H, Tintoré M, Montalban X. Altered maturation of circulating dendritic cells in primary progressive MS patients. *J Neuroimmunol.* 2006;175(1-2):183-191.
38. Martin R, McFarland HF, Boggs JM. Immunological aspects of experimental allergic encephalomyelitis and multiple sclerosis. *Crit Rev Clin Lab Sci.* 1995;32(2):121-182.
39. Hawiger D, Inaba K, Dorsett Y, et al. Dendritic cells induce peripheral T cell unresponsiveness under steady state conditions in vivo. *J Exp Med.* 2001;194(6):769-780.
40. Schwab N, Zozulya AL, Kieseier BC, Toyka KV, Wiendl H. An imbalance of two functionally and phenotypically different subsets of plasmacytoid dendritic cells characterizes the dysfunctional immune regulation in multiple sclerosis. *J Immunol.* 2010;184(9):5368-5374.
41. Ganguly D, Haak S, Sisirak V, Reizis B. The role of dendritic cells in autoimmunity. *Nat Rev Immunol.* 2013;13(8):566-577.
42. Pashenkov M, Huang Y-M, Kostulas V, Haglund M, Söderström M, Link H. Two subsets of dendritic cells are

- present in human cerebrospinal fluid. *Brain*. 2001;124(3):480-492.
43. Zang YC, Skinner SM, Robinson RR, et al. Regulation of differentiation and functional properties of monocytes and monocyte-derived dendritic cells by interferon beta in multiple sclerosis. *Mult Scler J*. 2004;10(5):499-506.
 44. Schreiner B, Mitsdoerffer M, Kieseier BC, et al. Interferon- β enhances monocyte and dendritic cell expression of B7-H1 (PD-L1), a strong inhibitor of autologous T-cell activation: relevance for the immune modulatory effect in multiple sclerosis. *J Neuroimmunol*. 2004;155(1-2):172-182.
 45. Vieira PL, Heystek HC, Wormmeester J, Wierenga EA, Kapsenberg ML. Glatiramer acetate (copolymer-1, copaxone) promotes Th2 cell development and increased IL-10 production through modulation of dendritic cells. *J Immunol*. 2003;170(9):4483-4488.
 46. Hu Z, Jiao Q, Ding J, et al. Berberine induces dendritic cell apoptosis and has therapeutic potential for rheumatoid arthritis. *Arthritis Rheum*. 2011;63(4):949-959.
 47. Yang Y, Qi J, Wang Q, et al. Berberine suppresses Th17 and dendritic cell responses. *Invest Ophthalmol Visual Sci*. 2013;54(4):2516-2522. doi:10.1167/iov.12-11217
 48. Karimi G, Mahmoudi M, Balali-Mood M, et al. Decreased levels of spleen tissue CD4+ CD25+ Foxp3+ regulatory T lymphocytes in mice exposed to berberine. *J Acupunct Meridian Stud*. 2017;10(2):109-113.
 49. Sallusto F. Heterogeneity of human CD4(+) T cells against microbes. *Annu Rev Immunol*. 2016;34:317-334. doi:10.1146/annurev-immunol-032414-112056
 50. Damsker JM, Hansen AM, Caspi RR. Th1 and Th17 cells: adversaries and collaborators. *Ann N Y Acad Sci*. 2010;1183:211-221. doi:10.1111/j.1749-6632.2009.05133.x
 51. Annunziato F, Cosmi L, Liotta F, Maggi E, Romagnani S. Human Th1 dichotomy: origin, phenotype and biologic activities. *Immunology*. 2014;144:343-351. doi:10.1111/imm.12399
 52. Szabo SJ, Kim ST, Costa GL, Zhang X, Fathman CG, Glimcher LH. A novel transcription factor, T-bet, directs Th1 lineage commitment. *Cell*. 2000;100(6):655-669. doi:10.1016/S0092-8674(00)80702-3
 53. Hsieh CS, Macatonia SE, Tripp CS, Wolf SF, O'Garra A, Murphy KM. Development of TH1 CD4+ T cells through IL-12 produced by Listeria-induced macrophages. *Science*. 1993;260(5107):547-549. doi:10.1126/science.8097338
 54. Kaskow BJ, Baecher-Allan C. Effector T cells in multiple sclerosis. *Cold Spring Harbor Perspect Med*. 2018;8(4):a029025. doi:10.1101/cshperspect.a029025
 55. Raphael I, Nalawade S, Eagar TN, Forsthuber TG. T cell subsets and their signature cytokines in autoimmune and inflammatory diseases. *Cytokine*. Jul 2015;74(1):5-17. doi:10.1016/j.cyto.2014.09.011
 56. Kunkl M, Frasca S, Amormino C, Volpe E, Tuosto L. T helper cells: the modulators of inflammation in multiple sclerosis. *Cells*. 2020;9(2):482. doi:10.3390/cells9020482
 57. Dhib-Jalbut S. Pathogenesis of myelin/oligodendrocyte damage in multiple sclerosis. *Neurology*. 2007;68(22 Suppl 3):S13-S21; discussion S43-S54. doi:10.1212/01.wnl.0000275228.13012.7b
 58. Raphael I, Forsthuber TG. Stability of T-cell lineages in autoimmune diseases. *Expert Rev Clin Immunol*. 2012;8(4):299-301. doi:10.1586/eci.12.22
 59. Skurkovich S, Skurkovich B. Anticytokine therapy, especially anti-interferon- γ , as a pathogenetic treatment in TH-1 autoimmune diseases. *Ann N Y Acad Sci*. 2005;1051:684-700. doi:10.1196/annals.1361.113
 60. de Weerd NA, Nguyen T. The interferons and their receptors—distribution and regulation. *Immunol Cell Biol*. 2012;90(5):483-491. doi:10.1038/icb.2012.9
 61. Hohnoki K, Inoue A, Koh CS. Elevated serum levels of IFN- γ , IL-4 and TNF- α /unelevated serum levels of IL-10 in patients with demyelinating diseases during the acute stage. *J Neuroimmunol*. 1998;87(1-2):27-32. doi:10.1016/S0165-5728(98)00053-8
 62. Lock C, Hermans G, Pedotti R, et al. Gene-microarray analysis of multiple sclerosis lesions yields new targets validated in autoimmune encephalomyelitis. *Nat Med*. 2002;8(5):500-508. doi:10.1038/nm0502-500
 63. Goris A, Heggarty S, Marrosu MG, Graham C, Billiau A, Vandebroek K. Linkage disequilibrium analysis of chromosome 12q14-15 in multiple sclerosis: delineation of a 118-kb interval around interferon- γ (IFNG) that is involved in male versus female differential susceptibility. *Genes Immun*. 2002;3(8):470-476. doi:10.1038/sj.gene.6363913
 64. Kantarci OH, Goris A, Hebrink DD, et al. IFNG polymorphisms are associated with gender differences in susceptibility to multiple sclerosis. *Gene Immun*. 2005;6(2):153-161. doi:10.1038/sj.gene.6364164
 65. Murphy AC, Lalor SJ, Lynch MA, Mills KHG. Infiltration of Th1 and Th17 cells and activation of microglia in the CNS during the course of experimental autoimmune encephalomyelitis. *Brain Behav Immun*. 2010;24(4):641-651. doi:10.1016/j.bbi.2010.01.014
 66. Prajeeth CK, Löhr K, Floess S, et al. Effector molecules released by Th1 but not Th17 cells drive an M1 response in microglia. *Brain Behav Immun*. 2014;37:248-259. doi:10.1016/j.bbi.2014.01.001
 67. Thompson AJ, Baranzini SE, Geurts J, Hemmer B, Ciccarelli O. Multiple sclerosis. *Lancet*. 2018;391(10130):1622-1636. doi:10.1016/S0140-6736(18)30481-1
 68. Ferber IA, Brocke S, Taylor-Edwards C, et al. Mice with a disrupted IFN-gamma gene are susceptible to the induction of experimental autoimmune encephalomyelitis (EAE). *J Immunol*. 1996;156(1):5-7.
 69. Willenborg DO, Fordham S, Bernard CC, Cowden WB, Ramshaw IA. IFN-gamma plays a critical down-regulatory role in the induction and effector phase of myelin oligodendrocyte glycoprotein-induced autoimmune encephalomyelitis. *J Immunol*. 1996;157(8):3223-3227.
 70. Ni C, Wang C, Zhang J, et al. Interferon- γ safeguards blood-brain barrier during experimental autoimmune encephalomyelitis. *Am J Pathol*. 2014;184(12):3308-3320. doi:10.1016/j.ajpath.2014.08.019
 71. Penberthy W, Tsunoda I. The importance of NAD in multiple sclerosis. *Curr Pharm Des*. 2009;15(1):64-99. doi:10.2174/138161209787185751
 72. Munn DH, Shafizadeh E, Attwood JT, Bondarev I, Pashine A, Mellor AL. Inhibition of T cell proliferation by macrophage tryptophan catabolism. *J Exp Med*. 1999;189(9):1363-1372. doi:10.1084/jem.189.9.1363
 73. Munn DH, Sharma MD, Mellor AL. Ligation of B7-1/B7-2 by human CD4+ T cells triggers indoleamine 2,3-dioxygenase

- activity in dendritic cells. *J Immunol.* 2004;172(7):4100-4110. doi:10.4049/jimmunol.172.7.4100
74. Liu X, Zhang X, Ye L, Yuan H. Protective mechanisms of berberine against experimental autoimmune myocarditis in a rat model. *Biomed Pharmacother.* 2016;79:222-230. doi:10.1016/j.biopha.2016.02.015
75. Qin X, Guo BT, Wan B, et al. Regulation of Th1 and Th17 cell differentiation and amelioration of experimental autoimmune encephalomyelitis by natural product compound berberine. *J Immunol.* 2010;185(3):1855-1863. doi:10.4049/jimmunol.0903853
76. Tong B, Yuan X, Dou Y, et al. Norisoboldine, an isoquinoline alkaloid, acts as an aryl hydrocarbon receptor ligand to induce intestinal Treg cells and thereby attenuate arthritis. *Int J Biochem Cell Biol.* 2016;75:63-73. doi:10.1016/j.biocel.2016.03.014
77. Li H, Li XL, Zhang M, et al. Berberine ameliorates experimental autoimmune neuritis by suppressing both cellular and humoral immunity. *Scand J Immunol.* 2014;79(1):12-19. doi:10.1111/sji.12123
78. Kang BY, Chung SW, Cho D, Kim TS. Involvement of p38 mitogen-activated protein kinase in the induction of interleukin-12 p40 production in mouse macrophages by berberine, a benzodioxoloquinolizine alkaloid. *Biochem Pharmacol.* 2002;63(10):1901-1910. doi:10.1016/s0006-2952(02)00982-6
79. Kips JC, Brusselle GJ, Joos GF, et al. Interleukin-12 inhibits antigen-induced airway hyperresponsiveness in mice. *Am J Respir Crit Care Med.* 1996;153(2):535-539. doi:10.1164/ajrccm.153.2.8564093
80. Shan K, Pang R, Zhao C, et al. IL-17-triggered down-regulation of miR-497 results in high HIF-1 α expression and consequent IL-1 β and IL-6 production by astrocytes in EAE mice. *Cell Mol Immunol.* 2017;14(11):909-923. doi:10.1038/cmi.2017.12
81. Ivanov II, McKenzie BS, Zhou L, et al. The orphan nuclear receptor ROR γ t directs the differentiation program of proinflammatory IL-17+ T helper cells. *Cell.* 2006;126(6):1121-1133. doi:10.1016/j.cell.2006.07.035
82. Saresella M, Tortorella P, Marventano I, et al. TH17-driven inflammation is present in all clinical forms of multiple sclerosis; disease quiescence is associated with Gata3-expressing cells. *Eur J Inflamm.* 2013;11:223-235. doi:10.1177/1721727X1301100121
83. Zambrano-Zaragoza JF, Romo-Martínez EJ, Durán-Avelar MJ, García-Magallanes N, Vibanco-Pérez N. Th17 cells in autoimmune and infectious diseases. *Int J Inflamm.* 2014;2014:1-12. doi:10.1155/2014/651503
84. Korn T, Bettelli E, Oukka M, Kuchroo VK. IL-17 and Th17 cells. *Annu Rev Immunol.* 2009;27:485-517. doi:10.1146/annurev.immunol.021908.132710
85. Kalra S, Lowndes C, Durant L, et al. Th17 cells increase in RRMS as well as in SPMS, whereas various other phenotypes of Th17 increase in RRMS only. *Mult Scler J Exp Transl Clin.* 2020;6(1):2055217319899695. doi:10.1177/2055217319899695
86. Komiyama Y, Nakae S, Matsuki T, et al. IL-17 plays an important role in the development of experimental autoimmune encephalomyelitis. *J Immunol.* 2006;177(1):566-573. doi:10.4049/jimmunol.177.1.566
87. Soltanmohammadi A, Tavaf MJ, Zargarani S, et al. Daphnetin alleviates experimental autoimmune encephalomyelitis by suppressing Th1 and Th17 cells and upregulating Th2 and regulatory T cells. *Acta Neurobiol Exp.* 2022;82(3):273-283. doi:10.55782/ane-2022-026
88. Durelli L, Conti L, Clerico M, et al. T-helper 17 cells expand in multiple sclerosis and are inhibited by interferon- β . *Ann Neurol.* 2009;65(5):499-509. doi:10.1002/ana.21652
89. Kebir H, Kreymborg K, Ifergan I, et al. Human TH17 lymphocytes promote blood-brain barrier disruption and central nervous system inflammation. *Nat Med.* 2007;13(10):1173-1175. doi:10.1038/nm1651
90. Prajeeth CK, Kronisch J, Khorooshi R, et al. Effectors of Th1 and Th17 cells act on astrocytes and augment their neuroinflammatory properties. *J Neuroinflamm.* 2017;14(1):204. doi:10.1186/s12974-017-0978-3
91. Fernando V, Omura S, Sato F, et al. Regulation of an autoimmune model for multiple sclerosis in Th2-biased GATA3 transgenic mice. *Int J Mol Sci.* 2014;15(2):1700-1718.
92. Noack M, Miossec P. Th17 and regulatory T cell balance in autoimmune and inflammatory diseases. *Autoimmun Rev.* 2014;13(6):668-677.
93. Yue M, Xia Y, Shi C, et al. Berberine ameliorates collagen-induced arthritis in rats by suppressing Th17 cell responses via inducing cortistatin in the gut. *FEBS J.* 2017;284(17):2786-2801. doi:10.1111/febs.14147
94. Raphael I, Nalawade S, Eagar T, Forsthuber T. T cell subsets and their signature cytokines in autoimmune and inflammatory diseases. *Cytokine.* 2015;74:5-17. doi:10.1016/j.cyto.2014.09.011
95. Xu X, Wang R, Su Q, et al. Expression of Th1- Th2- and Th17-associated cytokines in laryngeal carcinoma. *Oncol Lett.* 2016;12(3):1941-1948. doi:10.3892/ol.2016.4854
96. Jäger A, Kuchroo VK. Effector and regulatory T-cell subsets in autoimmunity and tissue inflammation. *Scand J Immunol.* 2010;72(3):173-184. doi:10.1111/j.1365-3083.2010.02432.x
97. Oreja-Guevara C, Ramos-Cejudo J, Aroeira LS, Chamorro B, Diez-Tejedor E. TH1/TH2 cytokine profile in relapsing-remitting multiple sclerosis patients treated with glatiramer acetate or natalizumab. *BMC Neurol.* 2012;12:95.
98. Steinman L, Conlon P. Antigen specific immunotherapy of multiple sclerosis. *J Clin Immunol.* 2001;21(2):93-98. doi:10.1023/a:1011020225433
99. Finkelman FD, Urban Jr, JF. The other side of the coin: the protective role of the TH2 cytokines. *J Allergy Clin Immunol.* 2001;107(5):772-780. doi:10.1067/mai.2001.114989
100. Haghmorad D, Yazdanpanah E, Sadighimoghaddam B, et al. Kombucha ameliorates experimental autoimmune encephalomyelitis through activation of Treg and Th2 cells. *Acta Neurol Belg.* 2021;121(6):1685-1692. doi:10.1007/s13760-020-01475-3
101. Kuchroo VK, Prabhu Das M, Brown JA, et al. B7-1 and B7-2 costimulatory molecules activate differentially the Th1/Th2 developmental pathways: application to autoimmune disease therapy. *Cell.* 1995;80(5):707-718. doi:10.1016/0092-8674(95)90349-6
102. Furlan R, Poliani P, Marconi P, et al. Central nervous system gene therapy with interleukin-4 inhibits progression of ongoing relapsing-remitting autoimmune encephalomyelitis

- in Biozzi AB/H mice. *Gene Therapy*. 2001;8(1):13-19. doi:10.1038/sj.gt.3301357
103. Broberg EK, Salmi AA, Hukkanen V. IL-4 is the key regulator in herpes simplex virus-based gene therapy of BALB/c experimental autoimmune encephalomyelitis. *Neurosci Lett*. 2004;364(3):173-178. doi:10.1016/j.neulet.2004.04.059
 104. Kostic M, Stojanovic I, Marjanovic G, Zivkovic N, Cvetanovic A. Deleterious versus protective autoimmunity in multiple sclerosis. *Cell Immunol*. 2015;296(2):122-132. doi:10.1016/j.cellimm.2015.04.006
 105. Lin WC, Lin JY. Berberine down-regulates the Th1/Th2 cytokine gene expression ratio in mouse primary splenocytes in the absence or presence of lipopolysaccharide in a preventive manner. *Int Immunopharmacol*. 2011;11(12):1984-1990. doi:10.1016/j.intimp.2011.08.008
 106. Ben-Nun A, Kaushansky N, Kawakami N, et al. From classic to spontaneous and humanized models of multiple sclerosis: impact on understanding pathogenesis and drug development. *J Autoimmun*. 2014;54:33-50. doi:10.1016/j.jaut.2014.06.004
 107. Viglietta V, Baecher-Allan C, Weiner HL, Hafler DA. Loss of functional suppression by CD4+CD25+ regulatory T cells in patients with multiple sclerosis. *J Exp Med*. 2004;199(7):971-979. doi:10.1084/jem.20031579
 108. Campbell DJ. Control of regulatory T cell migration, function, and homeostasis. *J Immunol*. 2015;195(6):2507-2513. doi:10.4049/jimmunol.1500801
 109. Fritzsche B, Haas J, König F, et al. Intracerebral human regulatory T cells: analysis of CD4+ CD25+ FOXP3+ T cells in brain lesions and cerebrospinal fluid of multiple sclerosis patients. *PLoS One*. 2011;6(3):e17988. doi:10.1371/journal.pone.0017988
 110. Almolda B, González B, Castellano B. Activated microglial cells acquire an immature dendritic cell phenotype and may terminate the immune response in an acute model of EAE. *J Neuroimmunol*. 2010;223(1-2):39-54. doi:10.1016/j.jneuroim.2010.03.021
 111. Yan Y, Zhang GX, Gran B, et al. IDO upregulates regulatory T cells via tryptophan catabolite and suppresses encephalitogenic T cell responses in experimental autoimmune encephalomyelitis. *J Immunol*. 2010;185(10):5953-5961. doi:10.4049/jimmunol.1001628
 112. Gärtner D, Hoff H, Gimsa U, Burmester GR, Brunner-Weinzierl MC. CD25 regulatory T cells determine secondary but not primary remission in EAE: impact on long-term disease progression. *J Neuroimmunol*. 2006;172(1-2):73-84. doi:10.1016/j.jneuroim.2005.11.003
 113. Zhang X, Reddy J, Ochi H, Frenkel D, Kuchroo VK, Weiner HL. Recovery from experimental allergic encephalomyelitis is TGF- β dependent and associated with increases in CD4+LAP+ and CD4+CD25+ T cells. *Int Immunol*. 2006;18(4):495-503. doi:10.1093/intimm/dxh390
 114. Buc M. Role of regulatory T cells in pathogenesis and biological therapy of multiple sclerosis. *Mediators Inflamm*. 2013;2013:1-11. doi:10.1155/2013/963748
 115. Haghmorad D, Yazdanpanah E, Jadid Tavaf M, et al. Prevention and treatment of experimental autoimmune encephalomyelitis induced mice with 1, 25-dihydroxyvitamin D3. *Neurol Res*. 2019;41:943-957. doi:10.1080/01616412.2019.1650218
 116. Zhang R, Zeng H, Zhang Y, et al. CD226 ligation protects against EAE by promoting IL-10 expression via regulation of CD4+ T cell differentiation. *Oncotarget*. 2016;7(15):19251-19264. doi:10.18632/oncotarget.7834
 117. Liu Y, Carlsson R, Comabella M, et al. FoxA1 directs the lineage and immunosuppressive properties of a novel regulatory T cell population in EAE and MS. *Nat Med*. 2014;20(3):272-282. doi:10.1038/nm.3485
 118. Beers DR, Henkel JS, Zhao W, et al. Endogenous regulatory T lymphocytes ameliorate amyotrophic lateral sclerosis in mice and correlate with disease progression in patients with amyotrophic lateral sclerosis. *Brain*. 2011;134(Pt 5):1293-1314. doi:10.1093/brain/awr074
 119. Reynolds AD, Banerjee R, Liu J, Gendelman HE, Lee Mosley R. Neuroprotective activities of CD4+CD25+ regulatory T cells in an animal model of Parkinson's disease. *J Leukoc Biol*. 2007;82(5):1083-1094. doi:10.1189/jlb.0507296
 120. Pedotti R, Farinotti M, Falcone C, et al. Allergy and multiple sclerosis: a population-based case-control study. *Mult Scler J*. 2009;15(8):899-906. doi:10.1177/1352458509106211
 121. Jiang Y, Wu A, Zhu C, et al. The protective effect of berberine against neuronal damage by inhibiting matrix metalloproteinase-9 and laminin degradation in experimental autoimmune encephalomyelitis. *Neurol Res*. 2013;35(4):360-368. doi:10.1179/1743132812Y.0000000156
 122. Ma X, Jiang Y, Wu A, et al. Berberine attenuates experimental autoimmune encephalomyelitis in C57 BL/6 mice. *PLoS One*. 2010;5(10):e13489. doi:10.1371/journal.pone.0013489
 123. Cui G, Qin X, Zhang Y, Gong Z, Ge B, Zang YQ. Berberine differentially modulates the activities of ERK, p38 MAPK, and JNK to suppress Th17 and Th1 T cell differentiation in type 1 diabetic mice. *J Biol Chem*. 2009;284(41):28420-28429. doi:10.1074/jbc.M109.012674
 124. Hu Z, Jiao Q, Ding J, et al. Berberine induces dendritic cell apoptosis and has therapeutic potential for rheumatoid arthritis. *Arthritis Rheum*. 2011;63(4):949-959. doi:10.1002/art.30202
 125. Li C, Xi Y, Li S, et al. Berberine ameliorates TNBS induced colitis by inhibiting inflammatory responses and Th1/Th17 differentiation. *Mol Immunol*. 2015;67(2 Pt B):444-454. doi:10.1016/j.molimm.2015.07.013
 126. Dardalhon V, Awasthi A, Kwon H, et al. IL-4 inhibits TGF- β -induced Foxp3+ T cells and, together with TGF- β , generates IL-9+ IL-10+ Foxp3- effector T cells. *Nature Immunol*. 2008;9(12):1347-1355. doi:10.1038/ni.1677
 127. Veldhoen M, Uyttenhove C, van Snick J, et al. Transforming growth factor- β 'reprograms' the differentiation of T helper 2 cells and promotes an interleukin 9-producing subset. *Nat Immunol*. 2008;9(12):1341-1346. doi:10.1038/ni.1659
 128. Matsushita T, Tateishi T, Isobe N, et al. Characteristic cerebrospinal fluid cytokine/chemokine profiles in neuro-myelitis optica, relapsing remitting or primary progressive multiple sclerosis. *PLoS One*. 2013;8(4):e61835. doi:10.1371/journal.pone.0061835
 129. Jäger A, Dardalhon V, Sobel RA, Bettelli E, Kuchroo VK. Th1, Th17, and Th9 effector cells induce experimental autoimmune encephalomyelitis with different pathological phenotypes. *J Immunol*. 2009;183(11):7169-7177. doi:10.4049/jimmunol.0901906

130. Ruocco G, Rossi S, Motta C, et al. T helper 9 cells induced by plasmacytoid dendritic cells regulate interleukin-17 in multiple sclerosis. *Clin Sci*. 2015;129(4):291-303. doi:10.1042/CS20140608
131. Li H, Nourbakhsh B, Ciric B, Zhang GX, Rostami A. Neutralization of IL-9 ameliorates experimental autoimmune encephalomyelitis by decreasing the effector T cell population. *J Immunol*. 2010;185(7):4095-4100. doi:10.4049/jimmunol.1000986
132. Nowak EC, Weaver CT, Turner H, et al. IL-9 as a mediator of Th17-driven inflammatory disease. *J Exp Med*. 2009;206(8):1653-1660. doi:10.1084/jem.20090246
133. Li H, Nourbakhsh B, Cullimore M, Zhang GX, Rostami A. IL-9 is important for T-cell activation and differentiation in autoimmune inflammation of the central nervous system. *Eur J Immunol*. 2011;41(8):2197-2206. doi:10.1002/eji.201041125
134. Trifari S, Kaplan CD, Tran EH, Crellin NK, Spits H. Identification of a human helper T cell population that has abundant production of interleukin 22 and is distinct from T(H)-17, T(H)1 and T(H)2 cells. *Nat Immunol*. 2009;10(8):864-871. doi:10.1038/ni.1770
135. Beyeen AD, Adzemovic MZ, Öckinger J, et al. IL-22RA2 associates with multiple sclerosis and macrophage effector mechanisms in experimental neuroinflammation. *J Immunol*. 2010;185(11):6883-6890. doi:10.4049/jimmunol.1001392
136. Xu W, Li R, Dai Y, et al. IL-22 secreting CD4+ T cells in the patients with neuromyelitis optica and multiple sclerosis. *J Neuroimmunol*. 2013;261(1-2):87-91. doi:10.1016/j.jneuroim.2013.04.021
137. Muls N, Nasr Z, Dang HA, Sindic C, van Pesch V. IL-22, GM-CSF and IL-17 in peripheral CD4+ T cell subpopulations during multiple sclerosis relapses and remission. impact of corticosteroid therapy. *PLoS One*. 2017;12(3):e0173780. doi:10.1371/journal.pone.0173780
138. Perriard G, Mathias A, Enz L, et al. Interleukin-22 is increased in multiple sclerosis patients and targets astrocytes. *J Neuroinflammation*. 2015;12:119. doi:10.1186/s12974-015-0335-3
139. Wing AC, Hygino J, Ferreira TB, et al. Interleukin-17- and interleukin-22-secreting myelin-specific CD4(+) T cells resistant to corticoids are related with active brain lesions in multiple sclerosis patients. *Immunology*. 2016;147(2):212-220. doi:10.1111/imm.12552
140. Almolda B, Costa M, Montoya M, González B, Castellano B. Increase in Th17 and T-reg lymphocytes and decrease of IL22 correlate with the recovery phase of acute EAE in rat. *PLoS One*. 2011;6(11):e27473. doi:10.1371/journal.pone.0027473
141. Kreyborg K, Eitzensperger R, Dumoutier L, et al. IL-22 is expressed by Th17 cells in an IL-23-dependent fashion, but not required for the development of autoimmune encephalomyelitis. *J Immunol*. 2007;179(12):8098-8104. doi:10.4049/jimmunol.179.12.8098
142. Wagner CA, Roqué PJ, Mileur TR, Liggitt D, Goverman JM. Myelin-specific CD8+ T cells exacerbate brain inflammation in CNS autoimmunity. *J Clin Invest*. 2020;130(1):203-213. doi:10.1172/JCI132531
143. Goldschmidt CH, Hua LH. Re-evaluating the use of IFN- β and relapsing multiple sclerosis: safety, efficacy and place in therapy. *Degener Neurol Neuromuscul Dis*. 2020;10:29-38. doi:10.2147/DNND.S224912
144. Sabatino Jr, JJ, Wilson MR, Calabresi PA, Hauser SL, Schneck JP, Zamvil SS. Anti-CD20 therapy depletes activated myelin-specific CD8(+) T cells in multiple sclerosis. *Proc Natl Acad Sci U S A*. 2019;116(51):25800-25807. doi:10.1073/pnas.1915309116
145. Tzartos JS, Friese MA, Craner MJ, et al. Interleukin-17 production in central nervous system-infiltrating T cells and glial cells is associated with active disease in multiple sclerosis. *Am J Pathol*. 2008;172(1):146-155. doi:10.2353/ajpath.2008.070690
146. Annibaldi V, Ristori G, Angelini DF, et al. CD161(high) CD8+T cells bear pathogenetic potential in multiple sclerosis. *Brain*. 2011;134(Pt 2):542-554. doi:10.1093/brain/awq354
147. Godfrey DI, MacDonald HR, Kronenberg M, Smyth MJ, Kaer LV. NKT cells: what's in a name. *Nat Rev Immunol*. 2004;4(3):231-237. doi:10.1038/nri1309
148. Brennan PJ, Brigl M, Brenner MB. Invariant natural killer T cells: an innate activation scheme linked to diverse effector functions. *Nat Rev Immunol*. 2013;13(2):101-117. doi:10.1038/nri3369
149. Godfrey DI, Stankovic S, Baxter AG. Raising the NKT cell family. *Nat Immunol*. 2010;11(3):197-206. doi:10.1038/ni.1841
150. Araki M. Th2 bias of CD4+ NKT cells derived from multiple sclerosis in remission. *Int Immunol*. 2003;15(2):279-288. doi:10.1093/intimm/dxg029
151. Illes Z, Kondo T, Newcombe J, Oka N, Tabira T, Yamamura T. Differential expression of NK T cell V alpha 24J alpha Q invariant TCR chain in the lesions of multiple sclerosis and chronic inflammatory demyelinating polyneuropathy. *J Immunol*. 2000;164(8):4375-4381. doi:10.4049/jimmunol.164.8.4375
152. O'Keefe J, Gately CM, Counihan T, et al. T-cells expressing natural killer (NK) receptors are altered in multiple sclerosis and responses to alpha-galactosylceramide are impaired. *J Neurol Sci*. 2008;275(1-2):22-28. doi:10.1016/j.jns.2008.07.007
153. De Biasi S, Simone AM, Nasi M, et al. iNKT cells in secondary progressive multiple sclerosis patients display pro-inflammatory profiles. *Front Immunol*. 2016;7:555. doi:10.3389/fimmu.2016.00555
154. Gausling R, Trollmo C, Hafler DA. Decreases in interleukin-4 secretion by invariant CD4-CD8-V α 24J α Q T cells in peripheral blood of patients with relapsing-remitting multiple sclerosis. *Clin Immunol*. 2001;98(1):11-17. doi:10.1006/clim.2000.4942
155. Gigli G, Caielli S, Cutuli D, Falcone M. Innate immunity modulates autoimmunity: type 1 interferon- β treatment in multiple sclerosis promotes growth and function of regulatory invariant natural killer T cells through dendritic cell maturation. *Immunology*. 2007;122(3):409-417. doi:10.1111/j.1365-2567.2007.02655.x
156. Mars LT, Laloux V, Goude K, et al. Cutting edge: V α 14-J α 281 NKT cells naturally regulate experimental autoimmune encephalomyelitis in nonobese diabetic mice. *J Immunol*. 2002;168(12):6007-6011. doi:10.4049/jimmunol.168.12.6007
157. Mars LT, Gautron AS, Novak J, et al. Invariant NKT cells regulate experimental autoimmune encephalomyelitis and infiltrate the central nervous system in a CD1d-independent manner. *J Immunol*. 2008;181(4):2321-2329. doi:10.4049/jimmunol.181.4.2321

158. Mars LT, Araujo L, Kerschen P, et al. Invariant NKT cells inhibit development of the Th17 lineage. *Proc Natl Acad Sci U S A*. 2009;106(15):6238-6243. doi:10.1073/pnas.0809317106
159. Hong S, Wilson MT, Serizawa I, et al. The natural killer T-cell ligand α -galactosylceramide prevents autoimmune diabetes in non-obese diabetic mice. *Nat Med*. 2001;7(9):1052-1056. doi:10.1038/nm0901-1052
160. Miyamoto K, Miyake S, Yamamura T. A synthetic glycolipid prevents autoimmune encephalomyelitis by inducing TH2 bias of natural killer T cells. *Nature*. 2001;413(6855):531-534. doi:10.1038/35097097
161. Horikoshi M, Goto D, Segawa S, et al. Activation of Invariant NKT cells with glycolipid ligand alpha-galactosylceramide ameliorates glucose-6-phosphate isomerase peptide-induced arthritis. *PLoS One*. 2012;7(12):e51215. doi:10.1371/journal.pone.0051215
162. Monteiro M, Almeida CF, Caridade M, et al. Identification of regulatory Foxp3+ invariant NKT cells induced by TGF- β . *J Immunol*. 2010;185(4):2157-2163. doi:10.4049/jimmunol.1000359
163. Kress E, Hedges J, Jutila M. Distinct gene expression in human V δ 1 and V δ 2 $\gamma\delta$ T cells following non-TCR agonist stimulation. *Mol Immunol*. 2006;43(12):2002-2011. doi:10.1016/j.molimm.2005.11.011
164. Stinissen P, Vandevyver C, Medaer R, et al. Increased frequency of gamma delta T cells in cerebrospinal fluid and peripheral blood of patients with multiple sclerosis. *J Immunol*. 1995;154(9):4883-4894.
165. Chen Z, Freedman MS. Correlation of specialized CD16(+) gammadelta T cells with disease course and severity in multiple sclerosis. *J Neuroimmunol*. 2008;194(1-2):147-152. doi:10.1016/j.jneuroim.2007.11.010

How to cite this article: Yazdanpanah E, Dadfar S, Shadab A, et al. Berberine: a natural modulator of immune cells in multiple sclerosis. *Immun Inflamm Dis*. 2024;12:e1213. doi:10.1002/iid3.1213